

**INTERNATIONAL JOURNAL OF ADVANCES IN
PHARMACY, BIOLOGY AND CHEMISTRY**

Research Article

**Ethnobotanical Study of Antitussive Plants Used in
Traditional Medicine by Abbey and Krobou
populations, in the South of Côte d'Ivoire**

N'Guessan Koffi*¹, Assi-Kaudjhis Chimène¹ and Kouassi K. Henri².

¹Université Félix Houphouët-Boigny (Côte d'Ivoire), U.F.R. Biosciences, Laboratoire de
Botanique; Institut Botanique Aké-Assi d'Andokoi (IBAAN)

²Université Jean Lorougnon Guédé (Côte d'Ivoire), U.F.R. Agroforesterie.

ABSTRACT

Cough is respiratory disease which becomes recidivism. Despite the development of antitussive medicines, it remains a major health problem. In the search of means of fighting, man used the medicinal properties of many plants. The ethnobotanical survey in Abbey and Krobou villages (Department of Agboville, Côte d'Ivoire), made it possible to discover that 40 traditional healers use 37 plants species for their antitussive effect. Drugs (leaf, flower, fruit, seed, root, rhizome, stem and barks) are used to develop 35 medicinal receipts. The monospecific receipts, 33 of them (94.28%), are mainly solicited. Leaves (40.54%) are more requested. The remedies are prepared by crushing, steaming, decoction, excision, expression, infusion, maceration, kneading, pounding, pulverization, softening and torrefaction. The preparation of the medicamentous receipts utilizes mainly decoction (19.04%). The majority of the remedies are employed by oral way, particularly out of drink (62.16%). *Oxyanthus unilocularis*, *Uapaca esculenta*, *Zanthoxylum gilletii*, *Kalanchoe crenata*, *Petersianthus macrocarpus*, *Iodes liberica*, *Spondias mombin*, *Picralima nitida* and *Triplochiton scleroxylon*, with over 75% efficacy indice, appear as the most efficient plants in cough treatment. The phytochemical screening shows that the antitussive effect would be the fact of various chemical compounds: alkaloids, catechin tannins, essential oils, flavonoids, mucilage, saponosides, polyphenols, sterols and polyterpenes.

Keywords: Agboville, Côte d'Ivoire, cough, Ethnobotany, Phytotherapy, traditional Healer

1. INTRODUCTION

Respiratory disease is a very serious problem that upset at certain times, humanity ¹. In worldwide, respiratory diseases are still among the leading causes of mortality, morbidity and consultation ². Cough, evidence symptom in various respiratory diseases, is treated in rural traditional areas, with plants. In Africa and in most of the developing countries, medicinal plants constitute the principal therapeutic arsenal available to healers ³. Despite the development of various medicines, cough remains a disease which is difficult to be eradicated. It is in this context that a genuine research activity on antitussive effects of plants was developed throughout Africa and

particularly in Côte d'Ivoire. In department of Divo, with Dida people, plants are used for their antitussive property ⁴. Various other ethnopharmacological studies have shown that African populations use plants to treat cough. We cite the works of Aké-Assi ⁵, in different areas of Africa and those of Nacoulma ⁶, in Burkina Faso. Unfortunately, in Africa, plants' properties are empirically appreciated. Thus, the chemical composition of drugs used daily by many people for health care is non known ⁷. This study is part an improvement and valorization program of traditional African medicine. To achieve an improvement of this African medicine, it is obvious

that a better knowledge of natural plant resources is required. This ethnobotanical study aims to discover more antitussive plants and provide scientific evidence of the effectiveness of their traditional effect using phytochemical and pharmacological characteristics.

2. MATERIAL AND METHODS

2.1. Study site

Located in Southern forest of Côte d'Ivoire, Agboville department is part of the guinea field of mesophilic sector, characterized by dense moist semi-deciduous forest⁸. Currently, the original vegetation has been degraded by human activities⁹. Annual average pluviometry is about 1400 mm of water. Its climate, warm and humid, is characterized by two seasons: a dry season from December to February and a long rainy season from March to November. Agboville population is estimated at 220050 souls¹⁰. Nearby native populations (Abbey and Krobou), there is a community of non-natives coming from other areas of Côte d'Ivoire and also a community of foreigners for the most part coming from West African sub-region.

2.2. Collection of plant material

A prospection was conducted in some villages of Agboville department, in the aim to research, in different ecological environments, plant species cited by the traditional healers and to take samples to build up a collection of dried plants. In laboratory, from collected samples, flora, various books and specimens of the herbarium of the National Floristic Center, we identified the plants species, by their scientific name and we determined their botanical characteristics.

2.3. Nomenclature

In this study, species nomenclature does not follow the traditional classification. Phylogenetic classification, according to the work of Angiosperms Phylogeny Group, in its latest version called APG III¹¹, was adopted, to name the listed plants. We followed the same principle regarding families, orders, clades, subphylums and phylums. The terms used to describe biological and phytogeographic types are borrowed to Aké-Assi¹².

2.4. Ethnopharmacological survey methodology

To conduct the survey on antitussive plants, various villages in Agboville department were visited. As approach, we met the healers (men and women) and organized semi-structured interviews. Each of them was visited twice, at different moments, to answer the same questions. That helped us to confirm or no the

informations we had already collected. During this ethnobotanical investigation, informations were collected relating to plants used in the treatment of cough, their therapeutic efficacy, the organs used as drugs, their methods of sampling and the modes of preparation and administration of medicamentous receipts. The informations we obtained from traditional healers, helped us to express the therapeutic efficacy of each plant species used in seeking the percentage of healed patients when 50 persons were treated. This approach has enabled us to establish therapeutic efficacy indices for listed plants species.

2.5. Phytochemical screening using experiences

To carry out the phytochemical screening, solvents (ether of oil, methanol and distilled water) and various classic reagents were used. The classical methods employed to characterize the chemical groups are described in the works of Békro *et al.*¹³ and N'Guessan *et al.*^{7,14}.

2.6. Non experimental validation for the medicinal activity of plants using phytochemical / pharmacological literature

According to the literature, we performed a validation of the traditional medical practices, by looking for the chemical constituents that explain the antitussive effect for the listed plants.

2.7. Treatment of data on ethnopharmacological survey

From informations collected on healed patients when 50 persons were treated, an efficacy indice expressed as a percentage (%) of each species of plant, was assigned as follows:

- in the range [01-05], the efficacy indice is estimated at 01-10
- in the range [06-10], the efficacy indice is estimated at 11-20
- in the range [11-15], the efficacy indice is estimated at 21-30
- in the range [16-20], the efficacy indice is estimated at 31-40
- in the range [21-25], the efficacy indice is estimated at 41-50
- in the range [26-30], the efficacy indice is estimated at 51-60
- in the range [31-35], the efficacy indice is estimated at 61-70.
- in the range [36-40], the efficacy indice is estimated at 71-80.
- in the range [41-45], the efficacy indice is estimated at 81-90.

-in the range [46-50], the efficacy indice is estimated at 91-100.

The plants, ranked according to their effectiveness, were subjected to a hierarchical classification, using STATISTICA software version 6.0, with Ward method.

3. RESULTS AND DISCUSSION

3.1. Botanical characteristics of the studied plants

The ethnobotanical investigations we conducted in Abbey and Krobou District (Department of Agboville, Côte d'Ivoire), made it possible to identify 37 species of antitussive plants, grouped in Angiosperms clade (table 1A). These species of plants belong to 37 genera and 24 families divided into 03 clades: Paleodicots (05 species), Monocots (03 species) and Eudicots (29 species), 01 subphylum (Tracheophytina) and 01 phylum (Embryophyta). The Dicots (Paleodicots and Eudicots), 34 of them, representing 91.89% of the identified species, have the highest number of plants. The predominance of Euphorbiaceae family (03 species or 8.10%), which has the largest number of medicinal plants used traditionally for their antitussive effects, can be explained by the fact that this family is part of the best represented in Ivorian flora¹². The comparison of our results with other works shows that inventories have the same characteristics in different studies. The ligneous species of taxonomic groups as Ferns and Gymnosperms have not been listed. Spermaphytes constitute the greater part of the therapeutic arsenal of antitussive plants. Our result is in accordance with previous reports, looking at the taxonomic groups of plants. For example, populations of Dakar, in Senegal, use 57 species of plants in the treatment of cough. All the listed plants belong to Spermaphytes. The percentage of Dicots (91.89%) we obtained approximates that of the Dicots (94.73%) in study conducted by Dioune (2006⁵), in Dakar (Senegal). However, variability was observed in the number of individuals identified in one study to another. We note, in study of Ouattara⁴, 09 species of plants representing 5.34% of the repertory of identified plants during ethnopharmacological study conducted in the area of Divo with Dida people, in South of Côte d'Ivoire. The Dicots have the highest number of plants. Aké-Assi⁵ undertook a significant study on the medicinal plants of Africa. However, only one plant: *Abrus precatorius* (Fabaceae) was mentioned like exerting antitussive effects. This variability would be due to the variations in the methods of investigation; it could be also explained by the differences of localities and vegetation.

3.2. Ethnopharmacological characteristics of listed plants

3.2.1. Visited sites and interviewed respondents

During this ethnopharmacological study relating to antitussive plants, 40 traditional healers, native of 10 villages in Agboville department, were met. They are men and women, but mostly men (75%) who agreed to collaborate with us by providing informations on antitussive plants. The oldest person of these healers is a man who is about 70 years and the youngest one is a woman of 30 years old. We met a lot of traditional healers who treat cough (08, or 20%) in Aboudé-Mandéké village.

3.2.2. Preparation and administration of herbal medicines

Various organs of the plant (leaf, flower, fruit, seed, root, rhizome, stem and barks) are used as drugs for medicinal preparations (table 1B). Leaves (40.54%) are the most requested organ. According to N'Guessan *et al.*¹⁵, the removal of 50% of plants leaves does not significantly affect the survival of this plant. Our result tallies with that of various studies conducted in other areas of Côte d'Ivoire and Africa^{4, 5, 6}. Also, the massive use of leaves would be the fact of their abundance. In terms of opotherapy, honey is necessary for the preparation of syrups. In the field of mineral therapy, the salt as seasoning is used, for example, in agreement with the statements of Fleurentin *et al.*¹⁶; authors indicated that, to relieve his pain and injuries, human uses its immediate environment. Various utensils are employed for the preparation of medicines: mortar, flat stone, pebble, canary, saucepan and calabash¹⁷. We distinguish various modes of preparation: crushing, steaming, decoction, excision, expression, infusion, maceration, kneading, pounding, pulverization, softening and torrefaction. Decoction (19.04%) is the most requested technique of preparation. The drug forms are diverse: decocte, exudates, extract, infuse, macerate, paste, powder and syrup. We note that 35 medicamentous receipts of remedies are developed to treat cough. The monospecific receipts, representing 94.28%, are mainly used. This result is similar to that of Ouattara⁴ who indicated that all the medicinal formulas used by Dida populations in Côte d'Ivoire, are monospecific. Considering the administration of medicines by oral routes, calabash, goblets, jugs, ladles, spoons, glasses (liqueur or beer) and cups are employed. The remedies are administered by absorption, drink, ingurgitation and oral instillations. The drink (62.16%) is the most widespread method of medicine administration. This result tallies with that of Ouattara⁴. In his study, author indicates that Dida of Divo (Côte d'Ivoire) use drink in 66.66% of cases.

3.2.3. Indices of efficacy

Indices of efficacy, determined on healed patients when 50 persons were treated, are shown in table 1B. A dendrogram (figure 1) represents the descriptive analysis. This analysis has established 04 groups of plants, when a cut is performed in the Euclidean distance 60. The first group contains plants with efficacy indice between 76 and 90%. We note 09 plants: *Iodes liberica*, *Kalanchoe crenata*, *Oxyanthus unilocularis*, *Petersianthus macrocarpus*, *Picralima nitida*, *Spondias mombin*, *Triplochiton scleroxylon*, *Uapaca esculenta* and *Zanthoxylum gillettii*. They have the best efficacy indice. These are excellent cough remedies. The second group has an efficacy indice between 61 and 75%. These plants are eleven (11): *Abelmoschus esculentus*, *Aframomum melegueta*, *Alstonia boonei*, *Annona muricata*, *Azadirachta indica*, *Boerhavia diffusa*, *Carapa procera*, *Cataranthus roseus*, *Ficus exasperata*, *Mangifera indica* and *Ocimum gratissimum*. They are good remedies against cough and are advisable in case of crisis. *Carica papaya*, *Chromolaena odorata*, *Cola nitida*, *Combretum paniculatum*, *Dissotis rotundifolia*, *Jatropha gossypifolia* and *Monodora myristica*, rank third in terms of efficacy indice which ranges from 46 to 60%; these are pretty-good remedies. The fourth group includes 10 species: *Citrus aurantifolia*, *Nymphaea lotus*, *Potomorphe guineense*, *Ricinodendron heudelotii*, *Scoparia dulcis*, *Terminalia catappa*, *Turnera ulmifolia*, *Xylopi aethiopica*, *Zea mays* and *Zingiber officinale*. They have an efficacy indice ranging from 34 to 45%. These are remedies that could be used if there is no other choice. We note that any plant is 100% effective; moreover, there is no plant with efficacy less than 34%.

3.2.4. Similarity with antitussive plants of other areas

Drugs sampling and preparation of herbal medicines vary according to ethnic groups, season, geographical and ecological environment of the traditional healers. Despite this variation, interesting similarities with some plants whose use is recognized in other ethnic groups were noticed. Decoction of *Mangifera indica* leaves is used by Abbey and Krobou populations in Agboville, to treat cough. This is also in agreement with the report of Jay (2014)¹⁸, on medicinal plants of his study. The stem barks of *Carapa procera* are employed against cough in Agboville department. These results share similarity to that of Diaouné (2006)⁵ who conducted ethnopharmacological investigations on antitussive plants in different markets of Dakar, in Senegal.

Carica papaya is employed for its antitussive properties, in Agboville. This result had also been reported on ethnobotanical study in Burkina Faso by Nacoulma⁶.

3.3. Phytochemistry

3.3.1. Experimental validation for antitussive effect of plants using phytochemistry

Primary validations of traditional medical practices, by looking for the chemical groups that explain the antitussive effects for some plants, were performed. Fourteen (14) plants of this study have been the subject of a phytochemical screening we have already carried out⁹. Table 2 gives the obtained results. The antitussive effect would be the fact of following chemical groups: alkaloids, catechics tannins, flavonoids, polyphenols, saponosides, sterols and polyterpenes (table 3).

3.3.2. Non experimental validation for antitussive effect of plants using phytochemical/ literature

The antitussive effect (table 4) results from several chemical elements: alkaloids, essential oils, flavonoids and mucilages. This phytochemical composition provides scientific evidence of the effectiveness of the traditional use of antitussive plants.

4. CONCLUSION

The ethnopharmacological investigations we conducted in Abbey and Krobou villages of Agboville (Côte d'Ivoire) show that 37 species of plants are used by 40 traditional healers, in various forms of medicinal preparations and administration, in the treatment of cough. Among the drugs, the leaves are mainly solicited. The preparation of the medicamentous receipts utilizes mainly decoction. The remedies are used particularly out of drink. The therapeutic effects are induced by various chemical compounds as alkaloids, catechics tannins, flavonoids, essential oils, mucilage, polyphenols, polyterpenes saponosides, sterols, which form the scientific basis of traditional therapeutic use of listed antitussive plants.

Table 1A
Taxonomic groups of plants used to treat cough in Abbey and Krobou villages

| Scientific names of plants species | Family | Clade |
|---|-----------------------|---------------------|
| <i>Abelmoschus esculentus</i> (L.) Moench | Malvaceae | Eudicot |
| <i>Aframomum melegueta</i> K. Schum. | Zingiberaceae | Monocot |
| <i>Alstonia boonei</i> De Wild. | Apocynaceae | Eudicot |
| <i>Annona muricata</i> L. | Annonaceae | Paleodicot |
| <i>Azadirachta indica</i> A. Juss. | Meliaceae | Eudicot |
| <i>Boerhavia diffusa</i> L. | Nyctaginaceae | Eudicot |
| <i>Carapa procera</i> DC. | Meliaceae | Eudicot |
| <i>Carica papaya</i> L. | Caricaceae | Eudicot |
| <i>Cataranthus roseus</i> (L.) G. Don | Apocynaceae | Eudicot |
| <i>Chromolaena odorata</i> (L.) King et H. Robins | Asteraceae | Eudicot |
| <i>Citrus aurantifolia</i> (Christm.) Swingle | Rutaceae | Eudicot |
| <i>Cola nitida</i> (Vent.) Schott et Endl. | Malvaceae | Eudicot |
| <i>Combretum paniculatum</i> Vent. | Combretaceae | Eudicot |
| <i>Dissotis rotundifolia</i> (Sm.) Triana | Melastomataceae | Eudicot |
| <i>Ficus exasperata</i> Vahl | Moraceae | Eudicot |
| <i>Iodes liberica</i> Stapf | Icacinaceae | Eudicot |
| <i>Jatropha gossypifolia</i> L. | Euphorbiaceae | Eudicot |
| <i>Kalanchoe crenata</i> (Andr.) Haw. | Crassulaceae | Eudicot |
| <i>Mangifera indica</i> L. | Anacardiaceae | Eudicot |
| <i>Monodora myristica</i> (Gaertn.) Dunal | Annonaceae | Paleodicot |
| <i>Nymphaea lotus</i> L. | Nymphaeaceae | Paleodicot |
| <i>Ocimum gratissimum</i> L. | Lamiaceae | Eudicot |
| <i>Oxyanthus unilocularis</i> Hiern | Rubiaceae | Eudicot |
| <i>Petersianthus macrocarpus</i> (P. Beauv.) Liben | Lecythidaceae | Eudicot |
| <i>Picralima nitida</i> (Stapf) T. Durand et H. Durand | Apocynaceae | Eudicot |
| <i>Pothomorphe guineense</i> Schum. et Thonn. | Piperaceae | Paleodicot |
| <i>Ricinodendron heudelotii</i> (Baill.) Pierre ex Pax | Euphorbiaceae | Eudicot |
| <i>Scoparia dulcis</i> L. | Plantaginaceae | Eudicot |
| <i>Spondias mombin</i> L. | Anacardiaceae | Eudicot |
| <i>Terminalia catappa</i> L. | Combretaceae | Eudicot |
| <i>Triplochiton scleroxylon</i> K. Schum. | Malvaceae | Eudicot |
| <i>Turnera ulmifolia</i> L. | Passifloraceae | Eudicot |
| <i>Uapaca esculenta</i> A. Chev. ex Aubrév. et Léandri | Euphorbiaceae | Eudicot |
| <i>Xylopia aethiopica</i> (Dunal) A. Rich. | Annonaceae | Paleodicot |
| <i>Zanthoxylum gillettii</i> (De Wild.) Wattermann | Rutaceae | Eudicot |
| <i>Zea mays</i> L. | Poaceae | Monocot |
| <i>Zingiber officinale</i> Rosc. | Zingiberaceae | Monocot |
| Total: Species: 37 Genera: 37 | Total of Families: 24 | Total of Clades: 03 |

Table 1B
Ethnobotanical characteristics of plants used to treat cough in Abbey and Krobou villages

| Scientific names of plants species | Organs used | Technic of preparation | Mode of administration | Therapeutic efficacy (%) |
|---|----------------|---|------------------------|--------------------------|
| <i>Abelmoschus esculentus</i> | Fruit, Root | Decoction: decocte | Drink | 60 |
| <i>Aframomum melegueta</i> | Seed | Mastication: extract | Ingurgitation | 62 |
| <i>Alstonia boonei</i> | Stem barks | Kneading: Paste + Palm oil | Absorption | 73 |
| <i>Annona muricata</i> | Flower | Infusion: Infusate | Drink | 64 |
| <i>Azadirachta indica</i> | Leaf | Sugary decoction: decocte | Drink | 72 |
| <i>Boerhavia diffusa</i> | Leaf | Kneading: Paste + Palm oil | Absorption | 66 |
| <i>Carapa procera</i> | Stem barks | Pounding: powder (maceration): macerate | Drink | 75 |
| <i>Carica papaya</i> | Root | Sugary decoction: decocte | Drink | 46 |
| <i>Cataranthus roseus</i> | Leaf | Sugary decoction: decocte | Drink | 68 |
| <i>Chromolaena odorata</i> | Leaf | Sugary decoction: decocte | Drink | 54 |
| <i>Citrus aurantifolia</i> | Fruit | Expression: Juice + honey (syrup) | Drink | 34 |
| <i>Cola nitida</i> | Seed | Crushing and maceration: macerate | Drink | 48 |
| <i>Combretum paniculatum</i> | Leaf | Kneading: Paste + water (mixture) | Drink | 60 |
| <i>Dissotis rotundifolia</i> ^{1*} | Leaf | Steamed | Oral instillations | 56 |
| <i>Ficus exasperata</i> ^{2*} | Stem barks | Kneading: Paste + Palm oil | Absorption | 70 |
| <i>Iodes liberica</i> ^{1*} | Leaf | Steamed | Oral instillations | 80 |
| <i>Jatropha gossypifolia</i> | Leaf | Decoction: decocte | Drink | 50 |
| <i>Kalanchoe crenata</i> | Leaf | Expression: juice + salt (syrup) | Drink | 82 |
| <i>Mangifera indica</i> | Leaf (joung) | Expression: juice + salt (syrup) | Oral instillations | 72 |
| <i>Monodora myristica</i> | Seed | Torrefaction, pulverization: powder + water | Drink | 54 |
| <i>Nymphaea lotus</i> | Leaf | Kneading: Paste + Palm oil | Absorption | 36 |
| <i>Ocimum gratissimum</i> | Leaf | Kneading: Paste + Palm oil | Absorption | 70 |
| <i>Oxyanthus unilocularis</i> | Stem barks | Mastication: extract | Ingurgitation | 90 |
| <i>Petersianthus macrocarpus</i> | Stem barks | Mastication: extract | Ingurgitation | 82 |
| <i>Picalima nitida</i> | Seed | Torrefaction, pulverization: powder + water | Drink | 76 |
| <i>Pothomorphe guineense</i> | Stem | Excision: exudate | Drink | 42 |
| <i>Ricinodendron heudelotii</i> | Stem barks | Decoction: decocte | Drink | 43 |
| <i>Scoparia dulcis</i> | Leaf | Softening, expression: extract | Drink | 44 |
| <i>Spondias mombin</i> | Leaf (joung) | Expression: juice + salt (syrup) | Oral instillations | 78 |
| <i>Terminalia catappa</i> | Stem barks | Softening, expression: extract | Drink | 42 |
| <i>Triplochiton scleroxylon</i> ^{2*} | Stem barks | Kneading: Paste + Palm oil | Absorption | 76 |
| <i>Turnera ulmifolia</i> | Flower | Infusion: infusate | Drink | 40 |
| <i>Uapaca esculenta</i> | Stem barks | Pounding: paste + palm oil | Absorption | 86 |
| <i>Xylopia aethiopica</i> | Fruit | Torrefaction, pulveisation: powder + water | Drink | 41 |
| <i>Zanthoxylum gillettii</i> | Leaf (joung) | Expression: juice + salt (syrup) | Oral instillations | 84 |
| <i>Zea mays</i> | Flower (style) | Decoction: decocte | Drink | 44 |
| <i>Zingiber officinale</i> | Rhizome | Mastication with salt: extract | Ingurgitation | 45 |

*: Bispecific receipts: ¹*Dissotis rotundifolia* and *Iodes liberica*; ²*Ficus exasperata* and *Triplochiton scleroxylon*

Table 2
Phytochemical screening of samples from 14 plant species

| Natural substances | | | Chemical groups | | | | | | | | | |
|----------------------------------|------------|------------|--------------------------|-------------|-------------|---------|----------|----------|-----------|-------------|-------------|---|
| Plants | Organs | Extracts | Sterols, polyterpenes | Polyphenols | Flavonoides | Tannins | | Quinones | Alkaloids | | Saponosides | |
| | | | | | | Gallic | Catechic | | Burchard | Dragendorff | | |
| <i>Aframomum melegueta</i> | Seed | Etheric | + | - | - | - | - | - | - | + | + | - |
| | | Methanolic | + | - | - | - | - | - | - | + | + | - |
| | | Aqueous | + | - | + | - | - | - | - | ++ | ++ | - |
| <i>Boerhavia diffusa</i> | Leaf | Etheric | + | - | - | - | - | - | - | + | + | - |
| | | Methanolic | + | - | - | - | - | - | - | + | + | - |
| | | Aqueous | - | + | + | - | - | - | - | + | + | + |
| <i>Chromolaena odorata</i> | Leaf | Etheric | - | - | + | - | - | - | - | + | + | - |
| | | Methanolic | - | + | + | - | - | - | - | + | + | - |
| | | Aqueous | + | + | + | - | + | - | - | + | + | + |
| <i>Cola nitida</i> | Stem barks | Etheric | + | - | + | - | - | - | - | - | - | - |
| | | Methanolic | + | + | + | - | - | - | - | - | - | - |
| | | Aqueous | + | + | + | - | - | - | - | - | - | + |
| <i>Ficus exasperata</i> | Leaf | Etheric | + | - | - | - | - | - | - | + | + | - |
| | | Methanolic | + | - | - | - | - | - | - | + | + | - |
| | | Aqueous | + | + | + | - | - | - | - | + | + | - |
| <i>Jatropha gossypifolia</i> | Stem barks | Etheric | + | - | - | - | - | - | - | + | + | + |
| | | Methanolic | + | - | - | - | - | - | - | + | + | + |
| | | Aqueous | + | - | - | - | + | - | - | + | + | + |
| <i>Mangifera indica</i> | Leaf | Etheric | + | + | + | - | - | - | - | + | + | - |
| | | Methanolic | + | + | + | - | + | - | - | + | + | - |
| | | Aqueous | + | + | + | - | + | - | - | + | + | + |
| <i>Monodora myristica</i> | Seed | Etheric | + | - | - | - | - | - | - | - | - | - |
| | | Methanolic | + | - | + | - | - | - | - | + | + | - |
| | | Aqueous | + | - | ++ | - | - | - | - | ++ | ++ | - |
| <i>Ocimum gratissimum</i> | Leaf | Etheric | + | - | - | - | - | - | - | + | + | - |
| | | Methanolic | + | + | - | - | - | - | - | + | + | - |
| | | Aqueous | + | + | + | - | + | - | - | ++ | ++ | - |
| <i>Petersianthus macrocarpus</i> | Stem barks | Etheric | + | + | + | - | - | - | - | + | + | - |
| | | Methanolic | + | + | + | - | - | - | - | + | + | - |
| | | Aqueous | - | + | + | - | + | - | - | + | + | + |
| <i>Potomorphe guineense</i> | Fruit | Etheric | + | - | + | - | - | - | - | + | + | - |
| | | Methanolic | + | - | + | - | - | - | - | + | + | - |
| | | Aqueous | + | - | ++ | - | - | - | - | ++ | ++ | - |
| <i>Terminalia catappa</i> | Stem barks | Etheric | - | - | + | - | - | - | - | - | - | - |
| | | Methanolic | - | + | + | - | - | - | - | + | + | - |
| | | Aqueous | - | + | + | - | - | - | - | + | + | + |
| <i>Xylopia aethiopica</i> | Fruit | Etheric | + | - | - | - | - | - | - | - | - | - |
| | | Methanolic | + | - | - | - | - | - | - | - | - | - |
| | | Aqueous | + | + | ++ | - | + | - | - | ++ | ++ | - |
| <i>Zingiber officinale</i> | Rhizome | Etheric | + | - | - | - | - | - | - | + | + | - |
| | | Methanolic | + | - | - | - | - | - | - | + | ++ | - |
| | | Aqueous | + | - | + | - | + | - | - | + | ++ | - |

Explanation of symbols: ++: Abundantly present; +: Presence; -: Absence

Table 3
Experimental validation for antitussive effect of plants using phytochemistry

| Plants species | Phytochemical screening (N'Guessan, 2008) ⁹ | Antitussive effect of plants using pharmacology | Literature |
|----------------------------------|--|---|---|
| <i>Aframomum melegueta</i> | Polyphenols | Immunostimulants providing resistance to aggression | (Betty, 2003) ¹⁹ |
| <i>Boerhavia diffusa</i> | Saponosides | Thinners, expectorant mucolytic action | (N'Guessan <i>et al.</i> , 2009) ⁷ |
| <i>Chromolaena odorata</i> | Polyphenols | Immunostimulants providing resistance to aggression | (Zirihi, 2006) ²⁰ |
| <i>Cola nitida</i> | Saponosides | Thinners, expectorant mucolytic action | (Fleurentin <i>et al.</i> , 2008) ²¹ |
| <i>Ficus exasperata</i> | Flavonoids | Bronchodilator, muscular relaxants | (Jay, 2014) ¹⁸ |
| <i>Jatropha curcas</i> | Saponosides | Thinners, expectorant mucolytic action | (Ahyi, 2001) ²² |
| <i>Jatropha gossypifolia</i> | Sterols and Polyterpenes | Antiseptic, disinfectant properties | (Fleurentin <i>et al.</i> , 2008) ²¹ |
| <i>Mangifera indica</i> | Flavonoids | Muscular relaxants | (Jay, 2014) ¹⁸ |
| <i>Monodora myristica</i> | Flavonoids | Muscular relaxants | (Ahyi, 2001) ²² |
| <i>Ocimum gratissimum</i> | Polysaccharides | Immunostimulants providing resistance to aggression | (Saraswathy, 2014) ² |
| <i>Petersianthus macrocarpus</i> | Tannins (catechics) | Bactericidal properties | (N'Guessan <i>et al.</i> , 2006) ²³ |
| <i>Potomorphe guineense</i> | Flavonoids | Muscular relaxants | (Jay, 2014) ¹⁸ |
| <i>Terminalia catappa</i> | Alkaloids | Vasodilators, they breake alveolar secretions | (Anonyme, 2015a) ²⁴ |
| <i>Xylopia aethiopica</i> | Flavonoids | Muscular relaxants | (Diaouné, 2006) ⁵ |
| <i>Zingiber officinale</i> | Flavonoids | Muscular relaxants | (Anonyme, 2015b) ²⁵ |

Table 4
Non experimental validation for antitussive effect of plants using phytochemical/ pharmacological literature

| Plants species | Phytochemistry | Pharmacology | Literature |
|---------------------------------|----------------|---|---|
| <i>Abelmoschus esculentus</i> | Mucilages | Emollient, protecting the pharyngeal mucosa from irritation | (Anonyme, 2015a) ²⁴ |
| <i>Alstonia boonei</i> | Alkaloids | Anti-cancer (tumors) | (Zirihi, 2006) ²⁰ |
| <i>Annona muricata</i> | Alkaloids | Respiratory stimulant | (Zirihi, 2006) ²⁰ |
| <i>Azadirachta indica</i> | Essential oils | Expectorant, bronchodilator, reduce phlegm | (Jay, 2014) ¹⁸ |
| <i>Carapa procera</i> | Flavonoids | Muscular relaxants | (Diaouné, 2006) ⁵ |
| <i>Carica papaya</i> | Alkaloids | Vasodilators, they brake alveolar secretions | (Diaouné, 2006) ⁵ |
| <i>Cataranthus roseus</i> | Alkaloids | Vasodilators, they brake alveolar secretions | (Anonyme, 2015a) ²⁴ |
| <i>Citrus aurantifolia</i> | Flavonoids | Muscular relaxants | (Diaouné, 2006) ⁵ |
| <i>Combretum paniculatum</i> | Alkaloids | Vasodilators, they brake alveolar secretions | (Osugwu and Nwoko, 2014) ²⁶ |
| <i>Dissotis rotundifolia</i> | Alkaloids | Vasodilators, they brake alveolar secretions | (Tavs <i>et al.</i> , 2010) ²⁷ |
| <i>Iodes liberica</i> | Alkaloids | Vasodilators, they brake alveolar secretions | (Fleurentin <i>et al.</i> , 2007) ¹⁶ |
| <i>Kalanchoe crenata</i> | Mucilages | Emollient, protecting the pharyngeal mucosa from irritation | (Jay, 2014) ¹⁸ |
| <i>Nymphaea lotus</i> | Alkaloids | excito-respiratory activity | (Tavs <i>et al.</i> , 2010) ²⁷ |
| <i>Picalima nitida</i> | Alkaloids | Vasodilators, they brake alveolar secretions | (Tavs <i>et al.</i> , 2010) ²⁷ |
| <i>Triplochiton scleroxylon</i> | Mucilages | Emollient, protecting the pharyngeal mucosa from irritation | (Fleurentin <i>et al.</i> , 2007) ¹⁶ |
| <i>Turnera ulmifolia</i> | Essential oils | Expectorant, bronchodilator, reduce phlegm | (Jay, 2014) ¹⁸ |
| <i>Zanthoxylum gillettii</i> | Alkaloids | Vasodilators, they brak alveolar secretions | (Zirihi, 2006) ²⁰ |
| <i>Zea mays</i> | Essential oils | Expectorant, bronchodilator, reduce phlegm | (Fleurentin <i>et al.</i> , 2008) ²¹ |

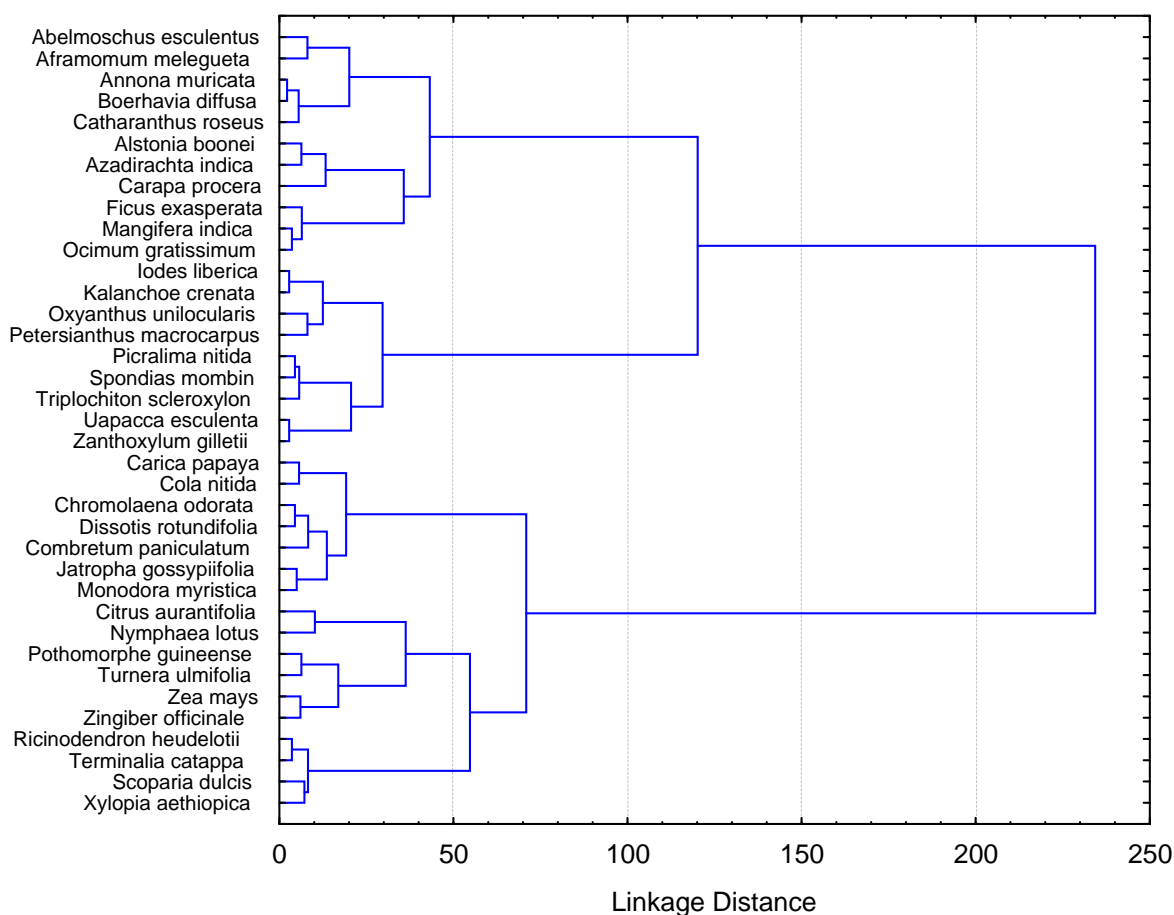


Figure 1
Dendrogram of efficacy indice for plants used in the preparation of cough remedies.

REFERENCES

1. Diaouné D. Plantes et médicaments utilisés contre la toux: enquête au niveau des marchés et des grossistes de la région de Dakar (Sénégal). Thèse pour obtenir le grade de Docteur en Pharmacie (Diplôme d'Etat). Université Cheikh Anta Diop de Dakar (Sénégal). Faculté de Médecine, de Pharmacie et d'Odonto-Stomatologie, 2006 ; 99 p
2. Saraswathy GR, Sathiya R, Anbu J, Maheswari E. Antitussive Medicinal Herbs - An Update Review. International Journal of Pharmaceutical Sciences and Drug Research, 2014; 6 (1): 12-19
3. Tahiya HAA, Amira HSAM, Mohammad AH, Afaf MW, Qasim AR. Comparative study of phytochemical screening, antioxidant and antimicrobial capacities of fresh and dry leaves crude plant extracts of *Datura metel*. Journal of King Saud University - Science, 2014; 6 (3): 237-243.
4. Ouattara D. Contribution à l'inventaire des plantes médicinales significatives utilisées dans la région de Divo (Sud forestier de la Côte d'Ivoire) et à la diagnose du poivrier de Guinée : *Xylopiia aethiopica* (Dunal) A. Rich. (Annonaceae). Thèse de Doctorat de l'Université de Cocody-Abidjan (Côte d'Ivoire), U.F.R. Biosciences, 2006 ; 184 p
5. Aké-Assi L. Abrégé de Médecine et pharmacopée africaines - Quelques plantes employées traditionnellement dans la couverture de soins de santé primaire, Edition NEI-CEDA, 2011 ; 157 p
6. Nacoulma-Ouédraogo O. Plantes médicinales et pratiques médicales traditionnelles au Burkina Faso : cas du Plateau central. Thèse de Doctorat ès Sciences Naturelles, Université de Ouagadougou (Burkina-Faso), Fac. Sc. et Tech., 1996 ; 605 p
7. N'Guessan K, Kadja B, Zirihi GN, Traoré D, Aké-Assi L. Screening phytochimique de quelques plantes médicinales ivoiriennes utilisées en pays Krobou (Agboville, Côte d'Ivoire). Sciences et Nature, 2009 ; 6 (1) : 1-15

8. Konaté S, Kampmann D. Atlas de la Biodiversité de l'Afrique de l'Ouest, Tome III : Côte d'Ivoire. Section 4 : Principaux facteurs environnementaux de la Côte d'Ivoire. Abidjan et Frankfurt/Main, 2010: 121-158.
9. N'Guessan K. Plantes médicinales et pratiques médicales traditionnelles chez les peuples Abbey et Krobou du Département d'Agboville (Côte d'Ivoire). Thèse de Doctorat d'Etat ès Sciences Naturelles, Spécialité Ethnobotanique, Université de Cocody-Abidjan (Côte d'Ivoire), U.F.R. Biosciences, Laboratoire de Botanique, 2008 ; 235 p
10. Koffi N, Kafana S, N'Guessan BYF. Étude anatomique de plantes utilisées en médecine traditionnelle en pays Abbey et Krobou, au Sud de la Côte-d'Ivoire. Int. J. Biol. Chem. Sci, 2012; 6 (1): 264-278
11. APG III (Angiosperm Phylogeny Group). Classification of the orders and families of flowering plants. Botanical Journal of the Linnean Society, 2009 ; 161 (2): 105-121
12. Aké-Assi L. Flore de la Côte d'Ivoire : catalogue systématique, biogéographique et écologique. Boissiera, Vol. 57, Conservatoire et Jardin Botanique de Genève, 2001 ; 396 p
13. Békro YA, Békro JAM, Boua BB, Tra-Bi FH, Éhilé EE. Etude ethnobotanique et screening phytochimique de *Caesalpinia benthamiana* (Baill.) Herend et Zarucchi (Caesalpinaceae). Rev. Sci. Nat., 2007; 4 (2): 217-225
14. N'Guessan K, Akédrin TN, Coulibaly K. Criblage phytochimique de plantes utilisées en médecine traditionnelle en pays Krobou (Côte d'Ivoire). Journal Africain de Communication Scientifique et Technologique, 2012 ; 14 : 1863-1877
15. N'Guessan K, Kouassi KH, Ouattara D. Plants used to treat anaemia, in traditional medicine, by Abbey and Krobou populations, in the South of Côte d'Ivoire. Journal of Applied Sciences Research, 2010 ; 6 (8): 1291-1297
16. Fleurentin J, Hayon JC, JM Pelt. Les plantes qui nous soignent-Traditions et thérapeutique. Editions Ouest-France, 2007 ; 189 p
17. N'Guessan K, Tra-Bi FH, Koné MW. Etude ethnopharmacologique de plantes antipaludiques utilisées en médecine traditionnelle chez les Abbey et Krobou d'Agboville (Côte d'Ivoire). Ethnopharmacologia, 2009; 44: 42-50
18. Jay RP. Antiasthmatic activity of traditional medicinal plants. International Journal of Pharmaceutical Sciences Review and Research, 2014; 5 (10):1000-1007
19. Betty JL. Plantes utilisées comme aphrodisiaques dans la réserve du Dja, Cameroun. Revue Med. Pharm. Afr., 2003 ; 17: 145-152
20. Zirih GN. Études botanique, pharmacologique et phytochimique de quelques plantes médicinales anti-paludiques et/ou immunogènes utilisées chez les Bété du Département d'Issia, dans l'ouest de la Côte d'Ivoire. Thèse de Doctorat d'Etat, Université de Cocody-Abidjan, U.F.R. Biosciences, 2006 ; 126 p
21. Fleurentin J, Hayon JC, JM Pelt. Plantes médicinales-Traditions et thérapeutique. Editions Ouest-France, 2008 ; 191 p
22. Ahyi RM. Le citronnier, cet autre arbre pharmacie. Revue Med. Pharm. Afr., 2001; 15: 113-119.
23. N'Guessan K, Kouassi KE, Zirih GN. Etude botanique et tri phytochimique de *Petersianthus macrocarpus* (P. Beauv.) Liben (Barringtoniaceae), une plante utilisée en médecine traditionnelle, dans la lutte contre le choléra. XII^{ème} colloque du Conseil Africain et Malgache pour l'Enseignement Supérieur (CAMES) ; Abidjan (Côte d'Ivoire) 2006. Pharmacopée et Médecine Traditionnelle Africaine, 2006 ; 14 : 125-143
24. <http://www.mon-erboristerie.com/blog/toux-et-phytotherapie/> (2015a)
25. <http://www.les-traitements.com/toux/tisanes-contre-la-toux.html> (2015b.)
26. Osuagwu GGE, Nwoko N. The phytochemical screening and antibacterial activity of the leaves of *Combretum paniculatum* (Vent), *Solanum macrocarpon* (L.) and *Catharanthus roseus* (L.) G. Don. J. of Pharmacy and Biological Sciences, 2014; 9 (1): 58-65
27. Tavs AA, Pius EO, Freddy OA. Antidiarrhoea and toxicological evaluation of the leaf extract of *Dissotis rotundifolia* Triana (Melastomataceae). BMC Complementary and Alternative Medicine, 2010; 10: 1-7