

**INTERNATIONAL JOURNAL OF ADVANCES IN
PHARMACY, BIOLOGY AND CHEMISTRY****Review Article****A review on Phytochemical analysis of
*Momordica charantia*****Prarthna Daniel, Ujjwala Supe*, M.G.Roymon.**

Plant Tissue Culture Laboratory, St. Thomas College, Bhilai,

District -Durg, Chattisgarh , India- 490006.

ABSTRACT

Momordica charantia commonly known as bitter melon/gourd, a member of Cucurbitaceae, is a slender, tendril climbing, annual vine. Bitter melon is a common food in tropics and is widely used as medicinal plants in countries like India, Brazil, China, Africa etc. In *M. charantia* primary metabolites are common sugars, proteins and chlorophyll while secondary metabolites are alkaloids, flavonoids, tanins, saponins, disogenin, proteins, calcium, copper etc. Secondary metabolites are responsible for medicinal activity of *Momordica charantia*. The medicinal values of Bitter melon lies in the bioactive phytochemical constituents that are non nutritive chemicals that produce definite physiological effects on human body and protect them from various diseases. Qualitative phytochemical analysis of *Momordica charantia* confirms the presence of phytochemicals like flavanoids, saponins, terpenoids, coumarins, emodins, alkaloids, proteins, cardiac glycosides, anthraquinones, anthocyanins, steroids etc.

Keywords : *Momordica charantia*, Regeneration, Phytochemical, callus.

INTRODUCTION

Momordica charantia L. commonly known as bitter melon/gourd, a member of Cucurbitaceae, is a slender, tendril climbing, annual vine. Bitter melon is a common food item of the tropics and is used for the treatment of cancer, diabetes and many ailments¹⁻⁴. It is a potent hypoglycemic agent^{5, 6} and hypoglycaemic actions for potential benefit in diabetes mellitus are possible due to at least three different groups of constituents in bitter melon. These include alkaloids, insulin like peptides, and a mixture of steroidal saponins known as charantin. Clinical studies with multiple controls have confirmed the benefit of bitter melon for diabetes⁷. Alpha and beta momarcharin are two proteins found in bitter melon, which are known to inhibit the AIDS virus⁸. *M. charantia* plant has not been much investigated for its *in vitro* culture response. However, formation of callus is reported⁹. *Momordica charantia* Linn. (bitter melon) belongs to the family of Cucurbitaceae. It is a climbing vine which is commonly seen growing on walls and shrubs in the tropics. The textured leaves look as a bite that is why the plant is given name Momordica which means to bite. The orange fruits are soft when ripe and have black seeds with a red covering.

**MEDICINAL PROPERTIES OF
MOMORDICA CHARANTIA**

Popularity of *Momordica charantia* in various system of traditional medicines for several ailments (antidiabetic, contraceptive, jaundice, abdominal pain, kidney (stone), piles, *pneumonia*, fever etc.) focused the investigator's attention on this plant¹⁰. Guanylate cyclase is thought to be linked to the pathogenesis and replication of not only psoriasis, but leukemia and cancer as well¹¹.

The anticancerous and antileukemic activity of bitter melon against numerous cell lines including liver cancer, human leukemia melanoma and solid sarcomas have also been documented¹². Saponins inhibit Na⁺ efflux leading to higher Na⁺ concentrations in cells, thereby activating a Na⁺, Ca²⁺ antiport. This effect produces elevated cytosolic Ca²⁺ which strengthens the contraction of the heart muscles and thereby reducing congestive heart failure¹³. Other uses include to expel intestinal gas, for tumors wound treatment, rheumatism, malaria, vaginal discharge and the seeds are used to induced abortion¹⁴. Bitter gourd has also been used as a traditional medicine for several other ailments, including dysentery, colic, fever, burns, painful menstruation, scabies and

other skin problems¹⁵. The presence of phenolic compounds in the plants indicates that these plants might have antimicrobial agent. Bitter melon might be used in the treatment of the placenta and navel of newborn baby which not only heals fast but also prevent the formation of infections^{16, 17}. The extract of crude bitter melon is used for different disease such as disease of liver and pancreas, anti-inflammatory, analgesic, reduces cholesterol level, promotes appetite and supports blood sugar management for diabetes of the people with high risk of developing diabetes¹⁸.

The plant extracts and juices have been found suitable for different diseases / problems¹⁹. Beside these stem and leaf of bitter melon is used in cancer treatment, in viral infections (HIV, herpes, Epstein Barr, hepatitis, influenza, and measles), in bacterial infections (Staphylococcus, Streptococcus, and Salmonella), as a bitter digestive aid (for dyspepsia and sluggish digestion) and in diabetes. The Bitter melon has many health benefits and medicinal properties. These are such as kills bacteria, reduce inflammation, kill viruses, fights free radicals, kills cancer cells, kills leukemia cells, prevents tumors, cleanses blood, reduces blood sugar and balance hormones¹⁴.

Phytochemical studies revealed plant to contain lutein and lycopene which are responsible for its antibiotic antitumor activities, charantin, momordicine and other alkaloids, saponins, phenolic constituents, glycosides and 5-hydroxytryptamine²⁰.

²¹ Plattel and Srinivasan, reported the hypoglycemic effect of the leaf extract of the plant. Antibacterial, antineoplastic, antiviral and antimutagenic activities of the plant have also been reported^{22, 23}. Sofowora, 1979 reported the purgative effect and the contractions of the guinea pig ileum of the plant extract. They also contain an array of biologically active proteins, namely, momordin, a- and b-momorcharin, cucurbitacin, and MAP30, that have shown to have highly effective anti-human immunodeficiency (HIV), anti-tumor anti-diabetic, and anti-rheumatic properties and to function as febrifuge medicine for jaundice, hepatitis, leprosy, hemorrhoids, psoriasis, snakebite, and vaginal discharge^{25,26,27}.

PHYTOCHEMICAL PROPERTIES:

The medicinal values of Bitter melon lies in the bioactive phytochemical constituents that are non nutritive chemicals that produce definite physiological effects on human body and protect them from various diseases. In *M.charantia* primary metabolites are common sugars, proteins and chlorophyll while secondary metabolites are alkaloids, flavonoids, tannins and so on (Table 1). Plant tissue culture have been investigated for industrial production of several useful secondary

metabolites^{28, 29}. Phenolic compounds, flavonoids are one of the widespread groups also acting as chemotaxonomic markers^{30, 31}. Phytosteroids are pharmacologically important for human life³². Diosgenin is synthesised in all plant parts with higher concentrations in fruit of *M.charantia*^{5, 33}. Exposure to salicylic acid frequently induces the synthesis of secondary metabolites in plant^{34, 35, 36}. Diosgenin is produced from different species of *Dioscorea*^{37, 38}. Flavonoids and steroids were carried out by using the methods of^{39, 40, 41, 42} respectively. Sterols are ubiquitous in higher plants and probably also in plant tissue cultures. *B-sitosterol* has been reported from *Nicotiana tabacum*⁴³, *M.charantia*⁹ and stigma sterol is reported from *Dioscorea tokoro*⁴² and *M.charantia*⁹. In plant pathogen interaction jasmonic acid and salicylic acid functions as endogenous signal compounds, thereby increasing the production of secondary metabolites by inducing the expression of defence related gene⁴⁴. Bitter melon has important role as a source of carbohydrate, protein, vitamins minerals and other nutrients in human diet⁴⁵. The presence of total sugar and starch was reported⁴⁶, reducing sugar⁴⁷, water soluble protein⁴⁸ and total protein by Micro – Kjeldahl method of AOAC⁴⁹. Vitamin C by the standard method of AOAC⁵⁰, lipid content⁵¹. Alkaloid with R.f.0.098 and 5-hydroxytryptamine is also reported⁵².

The protein content of bitter melon was reported⁵³, and protein of unpeeled bitter melon⁵⁴. The high amount of calcium and copper was reported in bitter melon^{53, 55}. charantin,⁵⁶ pure D-galacturoic acid⁵⁷, sterol glucoside⁵⁸ isolated a polypeptide –P all the phytochemicals possess hypoglycaemic action. Alkaloids and saponins are present in *Momordica* and volatile components are released during cooking which enhance the flavour^{59, 60}. Bitter melon plants contains high levels of iron, beta carotene, calcium, potassium, vitamins, phosphorus, and good dietary fiber^{61, 62}. *Xiao et al* found that the seed oil of *M.charantia* contained saturated fatty acids mainly steric acids, monounsaturated fatty acids like linoleic acid and polyunsaturated fatty acids.

In vitro regeneration of *Momordica charantia*

There are two ways to regenerate plants through the initiation of adventitious buds or through somatic embryos⁶⁴. In the present study, shoot and leaf explants were taken from aseptically grown seedling. tip, nodal and Internodal explants. A final treatment with 0.1% HgCl₂ was given for 5 mins after which explants were washed with distilled water. The explants were then inoculated on 15 ml aliquots of 0.8% agar containing MS medium⁶⁵ and incubated in dark for 20 days. When the radical emerged from the seeds, they were

transferred to incubation at $25 \pm 20^{\circ}\text{C}$ with a 16 h photoperiod provided by cool fluorescent light ($50 \mu \text{Em-2s-1}$) (Phillips, India). Approximately 2000-3000 lux artificial light intensity is needed. Cool white fluorescent light lamps are generally used for providing light. Generally 55 -60 % relative humidity is maintained in the culture room. After 5 weeks, from 15 cm long seedlings, 5-6mm shoot tip explants, 20-25 mm long nodal and internodal explants were dissected and inoculated on MS medium with various concentrations and combinations of growth regulators .

Cytokinin concentration of 0.5 mg/l to 6 mg/l produced shoots after 20 days in culture and the best response was observed on media containing 0.5 mg/l and 2 mg/l BAP, while IBA/NAA were suitable for rooting with best response at 4 mg/l IBA and 2mg/l NAA, the root formation was observed after 22 days in culture. Callus was formed on 2,4-D, with profuse callusing at 2mg/l of 2,4-D. A combination of NAA+BAP+2,4-D was most effective for callus formation with best response in 2mg/ l NAA + 0.5 mg / l BAP + 2 mg/l+2,4-D . This is in agreement that ^{66,67, 68} the tissue organ used as a source of explant can also be a determinant for the success of a plant tissue culture. Our results support the observation ⁶⁹ that organogenesis is determined by auxins and cytokinins. When tissues *in vitro* do not appear to require exogenous supply of auxins and cytokinin, it may be that sufficient endogenous levels of hormones exist in the culture system for organogenesis. MS medium without hormones also shows medium multiple shoots as well as roots formed.

The secondary metabolites are of immense importance for use as commercially as well as biologically active compounds. Flavonoids are naturally occurring phenolic compounds, which have a widespread distribution in intact plants and have been found in a number of tissue cultures. In callus cultures, the maximum amount of total steroid content was observed in six-week-old callus (27.34mg/gdw) and minimum was in 2 weeks old callus cultures (12.12mg/gdw) . The steroidal sapogenin, diosgenin is found in many plants but it is obtained principally from *Dioscorea* roots (4 to 6%dw) for conversion to commercially useful drugs. The presence of diosgenin was observed by thin layer chromatography coinciding with its standard sample in Rf values (Diosgenin-0.50). The color reaction test when sprayed with 50% H_2SO_4 fluorescent spot coinciding with their respective standard marker (Diosgenin-Green).

The liquid MS medium with NAA (2mg/l) + BAP (0.5mg/l) + 2, 4-D (2mg/l) was prepared. Callus was transferred to each flask and liquid cultures were grown on reciprocal shakers (125rpm; 5cm/stroke) for 6 days, on the 6th day liquid MS media was

supplied with three concentration (0.1mM, 0.05mM, 0.025mM) of salicylic acid and the cultures were again grown on same shakers for exactly 24h. The tissue grown on different media were harvested and weighted again and dried at room temperature. The callus samples were powdered and subjected to qualitative and quantitative estimation of diosgenin.

In vitro clonal propagation of *M.charantia* has been done ⁷⁰ leading to shoot and root differentiation at different level of cytokinin (BAP, Kin) and auxin (IBA, IAA, NAA) in MS medium. Leaf explants showed maximum callus percentage and callogenic response than other two explants stem and cotyledons. At different concentration of BAP and Kin green, compact and hard calluses produced. Best response towards multiple shoot regeneration was obtained from the nodal segments of *M.charantia* on MS medium supplemented with BAP and NAA .It was suggested by ⁷¹ that the combination of 2,4-D and BAP is most suitable for callus induction from shoots.

The shoots regenerated from callus, seedling explants and from multiple shoots were separated and transferred on MS medium containing different concentrations of rooting hormones, best response was on 3mg/l IBA, and the roots were developed roots in ten days . The complete plantlets thus formed, were hardened in green house and transferred to pots where 40% plants survived successfully. This is in agreement with previous report that multiple shoots can be formed on MS medium without hormones.

This study concludes that endogenous and exogenous level of growth regulators is also important for callogenesis and for differentiation.

Significance of *in-vitro* regeneration of *Momordica charantia*

The production of adventitious root and shoot from the cells of the tissue culture is called 'organogenesis'. If any wound or cut in plant is caused by any reason such as mechanical, chemical or by any infective agent they will form an unorganised proliferated mass of cell known as callus. When callus is grown aseptically on artificial nutrient medium in the glass vials under controlled experimental condition Genteret was first to successful promote the development of cell tissue. It is difficult and taking too much time to breed using normal method, while tissue culture is an efficient method for plant breeding and can shorten breeding period. The auxins are used to promote shoot growth and cytokinins are involved for root growth.

Due to valuable factors of *Momordica charantia* it is utilized as food and medicine importance in countries like China, India, Africa and West Indies since ancient times . Present study was conducted

for micropropagation using different explants through *in vitro* regeneration large number of plants can be produced through callus.

Tissue by manipulating hormonal and nutrient constituents in medium. The callus tissue from explants can provide genetic variability in *Momordica charantia*. Through callus culture of bitter gourd several secondary metabolites and biochemical assays can be produced⁷¹. All the

varieties were found to contain tannin, flavonoids, terpenoids, cardiac glycosides, triterpin and sterol, resin, amino acids and phenolic compounds except coumarin and free anthraquinones .

Plant tissue culture has been found to have potentials as a supplement to traditional agriculture in the industrial production of bioactive plant metabolites⁷².

Table1: Phytochemical analysis of Leaf and Callus extract

TESTS	LEAF EXTRACT	CALLUS EXTRACT
TANINS	+	+
ANTHOCYANINS	+	-
LEUCOANTHOCYANINS	-	-
FATTY ACIDS	-	-
COUMARINS	+	+
EMODINS	+	+
QUINONES	-	+
CARBOHYDRATES	-	-
CHOLESTROL	-	-
AMINO ACIDS	-	-
CARDIAC GLYCOSIDES	+	+
PHLOBATININS	-	-
<u>TEST FOR PHENOLS</u>		
FeCl ₂ TEST	+	-
LIEBERMANN' s TEST	+	-
<u>TEST FOR PROTEINS</u>		
NINHYDRIN TEST (aqueous)	-	-
NINHYDRIN REAGENT	+	+
OTHER TESTS	-	-
<u>TEST FOR ANTHRAQUINONES</u>		
BORNTAGER' s TEST	+	+
NINHYDRIN REAGENT	+	-
BIURET TEST	-	+
<u>TEST FOR ALKALOIDS</u>		
MAYER' s TEST	+	+
HAGER' s TEST	+	+
WAGNER' s TEST	-	-
OTHER TEST	+	+
<u>TEST FOR FLAVONOIDS</u>		
NaOH TEST	+	-
H ₂ SO ₄ TEST	-	+
OTHER TEST	+	+
<u>TEST FOR STEROLS</u>		
TERPENOIDS	+	+
SAPONINS	+	+

REFERENCES

1. Cefalu WT, Ye J, Wang ZQ, . Efficacy of dietary supplementation with botanicals on carbohydrate metabolism in humans. *endocrine, Metabolic & Immune disorders-Drug Targets* 2008 ; 8:78-81
2. Leung L, Birtwhistle R, Kotecha J, Hannah S, Cuthbertson S. Anti-diabetic and hypoglycaemic effects of *Momordica charantia* (bitter melon): A mini review, *British Journal of Nutrition*, (2009), (in press).
3. Modak M, Dixit P, Londhe J, Ghaskadbi S, . Indian herbs and herbal drugs used for the treatment of diabetes. *J. Clin. Biochemistry Nutri*, 2007; 40: 163- 173.
4. Nahas R, Moher . Complementary and alternative medicine for the treatment of type 2 diabetes. *Can Fam Physician* 2009; 55:591-596
5. Basch E, Gabardi S, Ulbaricht C. Bitter melon (*Momordica charantia*): A review of efficacy and safety. *Am J. Health Syst Pharm*, 2003; 60: 356-359.
6. Singh J, Cumming, E, Manmohan, G, Kalasz, H, Adeghate, E .The Open Medicinal Chemistry Journal 2011; 59:70-77.
7. Raman A, Lau C, Anti-diabetic properties and phytochemistry of *Momordica charantia* L. *Phytome* 1996; 349-62
8. Zhang QC, Preliminary report on the use of *M. charantia* extracted by HIV patients. *J. naturopath medicine* 1992; 3: 65-69.
9. Khanna P, Mohan S, Isolation and identification of diosgenin and sterols from fruits and in vitro cultures of *Momordica charantia* L., *Indian J. Exp. Biol.* 1973; 11: 58-60.
10. Grover, J.K. and S.P. Yadav. Pharmacological actions and potential uses of *Momordica charantia* . A review *J. Ethnopharmacol*, 2004; 93: 123-132.
11. Takemoto, D.J. Jilka C, Rockenbach S, Hughes JV. Purification and Characterization of a cytostatic factor with anti viral activity from the bitter melon. *Prep. Biochem.* 1983 ; 371-393.
12. Zhu, Z.J. Zhong ZC, Luo ZY, Xiao ZY. Studies on the active constituent of *Momordica charantia*. 1990; 25:898 -903.
13. Schneider and Wolfing, J. Synthetic cardenolids and related compounds. *Current organic chemistry* 2004 ; 8 No. 14.
14. Taylor ,L. Bitter melon: Herbal properties and Actions . In : *the Healing Power of Rainforest Herbs* , Taylor, L . (Ed) , Square One Publication Inc., New York 2005 pp 1-5.
15. Beloin N, Gbeassor M, Akpagana K, Hudson J, de Soussa K, Koumaglo K, Arnason JT. Ethanomedical uses of *Momordica charantia* (Cucurbitaceae) in Togo and relation to its phytochemistry and biological activity, *J. Ethnopharm*, 2005; 96: 45-55.
16. Okwu, D.E. Evaluation of the chemical composition of indigenous species and flavouring agents. *Global J. Pure Applied Sci* , 2001; 7: 455-459 .
17. Okwu, D.E.. The potentials of *Ocimum gratissimum*, *Pergularia extensa* and *Tetrapleura tetraptera* as spice and flavouring agents. *Nig. Agri. J.*, 2003; 35:143-148.
18. Verma, J.P. and J.S. Agarwal . Anote on the component fatty acid of the oil from the seeds of *Momordica charantia* L. *J. Indian Chem Soc* , 1956; 33: 355- 357.
19. Nadakarni, K. M. *Momordica charantia* L. *Indian. Material. Medica. Vol. I, part II*, 1982; pp. 805-807.
20. Dhalla NS, Gupta KC, Sastry MS and Malhotra CL . Chemical composition of the fruit of *Momordica charantia* Linn. *India J Pharm.*, 1981; 23: 128-131
21. Plattel, K., Srinivasan, K. Plant foods in the management of diabetes mellitus: vegetables as potential hypoglycemic agents. *Nahrung* , 1997; 41: 68-74.
22. Jilka C, Striffler B, Fortner GW, Hays EF, Takemoto DJ. . *In vivo* antitumor activity of the bitter melon (*Momordica charantia*). *Cancer Res.*, 1983; 443: 5151-5155
23. Guevara, A.P., Lim-sylianco, C., Cayrit, F., Finch, P. Anti-mutagens from *Momordica charantia*. *Mutat Res*, 1990; 230: 121-126
24. Sofowora A. Proceedings of a symposium on stigmatodiolenol from *Momordica charantia*. *Tetrahedron Lett.*, 1979; 26: 2217-2221.
25. Singh A, Singh SP, Bamezai R. *Momordica charantia* (Bitter gourd) peel, pulp, seed and whole fruit extract inhibits mouse skin papillomagenesis. *Toxicology Letters*, 1990 ; 94: 37-46.
26. Bourinbaiar AS, Lee-Huang S . The activity of plant derived antiretroviral proteins MAP30 and GAP31 against herpes simplex virus *in vitro*. *Biochemical and Biophysical Research Communication* 1996; 219: 923-929.
27. Beloin N, Gbeassor M, Akpagana K, Hudson J, Soussa KD, Koumaglo K, Arnason JT. Ethnomedicinal uses of *Momordica charantia* (Cucurbitaceae) in Togo and relation to its phytochemistry and biological activity. *J Ethnopharmacol* 2005; 96:49-55
28. Rao RS., Ravishankar GA. Plant Tissue cultures; Chemical factories of secondary metabolites. *Biotechnol. adv.*; 20: 101-153.

29. Wink et al , Sustainable biproduction of phytochemicals by plant *in vitro* cultures: anticancer agents . Plant Genetic Resources 2005; 3:90-100
30. Harborne JB, Current trends in chromatographic analysis of plant phenols, Adv. Med
31. . Shahidul Islam, Mohammad Jalaluddin , and Navam S. Hettiarachchy .Bioactive compounds of Bitter melon genotypes (*Momordica charantia* L.) in relation to their physiological functions.Fuctional foods in Health and Diseases 2011;1(2):61-74.
32. Hardman R, Recent advances in plants of pharmacological importance. Steroidal aspects, Egypt.J .Pharm.Sec.1980; 21 (12):169.
33. Taylor L, Tropical plant databases. Available at: <http://www.rain-tree.com /plants. htm>. Accessed 08/10/04.
34. Benhamou N. Elicitor-induced plant defense pathways. Trends in Plant Sciences. 1996; 1: 233-240.
35. Namdeo, AG. Plant cell elicitation for production of secondary metabolites; A review.Pharmacognosy Rev 2007; 1:69-79.
36. Jeong GT, Park DH. Enhanced secondary metabolites biosynthesis by elicitation in transformed plant root system.Appl.Biochem.biotechnol.130:436-446
37. Staba EJ, Milestones in plant tissue culture systems for production of secondary metabolites, J Nat Prod 1985; 48(2): 203-209.
38. Tomita Y, Uomori A, Structure and biosynthesis of Protookoronin steroidal sapogenins and sterols in tissue cultures of *Dioscorea tokoro*. Phytochemistry1974; 13:729.
39. Ozeki Y, Komamine J, Induction of antocynin synthesis in relation to embryogenesis in a carrot suspension culture, A model system for the study of expression and repression of plant cell cultures, In: Neumann KH, Barz W, Reinherd E. (Eds.) Primary and secondary metabolism of plant cell cultures Springer-Verlag, Berlin-Heidelberg, New York (1985) P 99.
40. Wong E, Francis CM, Flavonoids in genotypes of *Trifolium subterranean*, I, The normal flavonoids pattern of the geraldation variety. Phytochemistry1968;7: 2123-29.
41. Mabary J J, Markham KR, Thomas MB, The systematic identification of flavonoids. (1970), Springer-Verlag, Berlin-New York. Heidelberg
42. Tomita Y,Uomori A,Minto H, Steroidal sapogenins and sterols in tissue culture of *Dioscorea tokoro*,Phytochemistry1970; 9:111.
43. Beneveniste P, Hirth L, Oiuirsson G. La biosyntheses sterol dans les tissue de tabacco cultiven *in vitro* II. Particularity de la biosyntheses des phytosterols des tissue de tabac cultivates *in vitro*. Phytochemistry .1966; 5:45-68.
44. Tatiana C, Spollansky Sandra I Pitta-Alvarez and Giulietti M. Effect of jasmonic acid and aluminum on production of tropane alkaloids in hairy root cultures of *Brugmansia candida*, Electronic J. of Biotech2000; 3: 72-75
45. Ali et al . Characteristic of seed oils and nutritional composition of seeds from different varieties of *Momomrdica charantia* Linn. Cultivated in Bangladesh. Czech J.Food Sci. , 2008; 26: 275- 283.
46. Jayaraman J.Laboratory Manual in Biochemistry . Wiley Eastern Ltd., New Delhi , India , 1981;pp:53.
47. Miller. G.L.Use of Dinitrosalicylic acid reagent for determination of glucose .Anal Chem., 1972;31:426-428.
48. Oliver H. Lowry, Nira J. Rosebrough, A. Protein measurement with the folin phenol reagent .J. Biol.Chem., 1951; 193 :265 -273.
49. AOAC.Official and Tentative Methods of Analysis 9th Ed. , Association of Official Analytical Chemists ,Washington DC. 1960 ,pp 33
50. AOAC .Official Methods of Analysis , Association of Official Analytical Chemists 1995;Washington DC
51. Bligh , E.G.and W.J.Dyer . A rapid method for lipid extraction and purification .Can J.Biochem.Physiol,37:911- 917 .
52. Dhalla NS,Gupta KC,Sastry MS and Malhotra CL .Chemical composition of the fruit of *Momordica charantia* L.Ind.J.Pharmacol , 1961;23:128-128
53. Soomro, A.K. , Ansari ,K.A. Medicinal uses of Bitter gourd (*Momrdica charantia*), Hamdard Med 2005; 48:9-14.
54. Alvi Shahnaz,Khan K.M.,A. Munir,Sheikh and Mohammad Shahid . Effect of peeling and cooking on nutrients in vegetable.Pak j. Nutri , 2003; 2:189-191.
55. Lotikar , M.M and M.R. Rajamaro Rao . Note on Hypoglycemic principle (charantin) from the fruits of *Momordica charantia* . 1963; Biol. Abstr. 42(2) ;7748 and J.V.Bombay Sect.29(2):223-224.
56. Vasistha, S.K. , Vasistha , V.R.K. Rao .Thru Chem Abstr 1962; 12: 228-230
57. Sucrow W, . Sterol glycoside and new stigmasterol fro *Momordica charantia* . Tetrahedron Letter 1965;26: 2217-2221(Ger) (Thur Chem Abstr 1965.63:8444g)
58. Khanna P, Jain SC, Panagariya A, Dixit VP. . Hypoglycemic activity of polypeptide –P

- from a plant source of natural products J. Nat, 1981;Prod.44 (6) : 648 – 655.
59. Binder, R. G., Flath R. A. & Mon, T. R. . Volatile components of bittermelon . *J. Agr. Food Chem.*, 1989; **37**: 418-420
60. Schultes, R.E. Biodynamic cucurbits in the New World tropics. In: Biology and utilization of the Cucurbitaceae. D. M. Bates, R. W. Robinson, & C. Jeffrey (eds.),1990 ; pp. 307-317.
61. Paul A, Raychaudhuri SS. Medicinal uses and molecular identification of two *Momordica charantia* varieties – a review. *Electronic Journal of Biology*.2010; 6(2): 43-51.
62. Sultana RS, Bari Miah MA *In vitro* propagation of karalla (*Momordica charantia* L). *J Biol Sci* ,2003;3:1134–1139
63. Liu XR, Deng ZY, Fan YW, Li J, Liu ZH.. Mineral elements analysis of *Momordica charantia* seeds by ICP-AES and fatty acid profile identification of seed oil by GC-MS. *Guang Pu Xue Yu Guang Pu Fen Xi.*, 2010; 30(8): 2265-8.
64. Torrey JG, Morphogenesis in relation to chromosomal constitution in long term plant tissue cultures, *Plant physiology*1966;20:265.
65. Murashige T, Skoog F, A revised medium for rapid growth and bioassays with tobacco tissue cultures, *Physiology plantarum*, 1962;15:473-479
66. Cheng TY, Voqui TH, Regeneration of Douglas fir plantlets through tissue culture, *Science*1977;198: 306
67. Murashige T, Suppression of shoot formation in cultured tobacco cells by Gibberellic acid. *Science* 1961; 134: 280
68. Thrope TA, Patel KR, Clonal propagation, Adventitious buds In; Vasil, I K (Ed) *Cell Culture and Somatic Cell Genetics Of Plants Vol .1*,Academic Press, New York 1984; P49-60
69. Kim SG, Chang JR, Cha HC, Lee KW, Callus growth and plant regeneration from cotyledons in diverse cultures of Cucumber (*Cucumis sativus* L.), *Plant Cell, Tissue Org. Cult* 1988;12:67-74.
70. Agarwal M. and R. Kamal. *Invitro* clonal propagation of *Momordica charantia* L. *Indian J. Biotechnol.*,2004; 3: 426-430.
71. Ramachandra Rao, S. & Ravishankar, G. A. Biotransformation of protocatechuic aldehyde and caffeic acid to vanillin and capsaicin in freely suspended and immobilized cell cultures of *Capsicum frutescens*. *J. Biotechnol.*, 2000;76: 137-146.
72. Supe, U. Effect of growth regulator on callus induction of *Bryonia lacinosa*. *International journal of applied biotechnology and Biochemistry.*, 2011;1(4): 449-452.